

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/293798395>

The Risk of Prion Infection through Bovine Grafting Materials

Article in *Clinical Implant Dentistry and Related Research* · February 2016

DOI: 10.1111/cid.12391

CITATIONS

2

READS

476

3 authors:



Yeongsug Kim

Children's Hospital Los Angeles

2 PUBLICATIONS 24 CITATIONS

SEE PROFILE



Angel Rodriguez

Boston University

4 PUBLICATIONS 15 CITATIONS

SEE PROFILE



Hessam Nowzari

University of Southern California

94 PUBLICATIONS 2,419 CITATIONS

SEE PROFILE

The Risk of Prion Infection through Bovine Grafting Materials

Yeongsug Kim, DDS, MSD;* Angel Emmanuel Rodriguez, DDS;† Hessam Nowzari, DDS, PhD‡

ABSTRACT

Background: Bovine-derived grafting materials are frequently used in a variety of bone augmentation techniques. The aim of this paper is to assess the unique safety issue of bovine-derived grafting materials that is rarely addressed in dental literature: risk of bovine spongiform encephalopathy (BSE).

Methods: The validity of the current BSE diagnostic methods, surveillance and epidemiological trends in affected countries, and BSE infectivity in bovine bone before and after manufacturing processing were reviewed and analyzed.

Results: Prion screening has significant limits. Humans are not safe from the infection of prion disease of other species. Prions can and do break the species barrier. There is evidence there may be tens of thousands of infectious carriers in the western countries alone. This raises concern about the potential for perpetuation of infection via medical procedures.

Conclusion: The limited ability to screen prions within the animal genome, along with a long latency period to manifestation of the disease (1 to over 50 years) in infected patients, provides a framework for discussing possible long-term risks of the xenografts that are used so extensively in dentistry. We suggest abolishing the use of bovine bone.

KEY WORDS: cCJD, EU, FDA, IHC, PMCA, PK-sensitive, PrP^C, PrP^{Sc}, PrP 27–30, SAF, sCJD, WB

INTRODUCTION

Bovine bone was first introduced in surgery by Orell in 1934.¹ Xenografts have been hardly validated in orthopedics,² whereas bovine-derived bone graft materials are commonly used in dentistry as an alternative to autogenous tissue. The unique safety issue of using these materials is rarely addressed in dental literature and appears to be ignored by practitioners: risk of bovine spongiform encephalopathy (BSE) transmission from bovine xenograft. BSE is a type of transmissible spongiform encephalopathies (TSE) or prion diseases, a group of fatal neurodegenerative diseases affecting humans and a broad spectrum of animal species.³

Creutzfeldt–Jakob disease (CJD) was first described in the 1920s by Creutzfeldt and Jakob as human TSE with long periods of incubation. Sporadic CJD, the most common form of CJD, has been estimated to affect 1 to 1.5 persons per million annually around the world (Table 1). An epidemic variant of CJD appeared in the 1990s in the United Kingdom and public and scientific attention was attracted to the understanding of the propagation and pathogenesis.^{4–8}

Different types of prion diseases share common molecular mechanisms in which the misfolding of PrP^C leads to the pathologic version of the protein called PrP^{Sc}. The “protein-only” hypothesis suggests that once PrP^{Sc} is formed, it serves as a template to propagate this version of the protein, promoting the misfolding of PrP^C.^{9,10}

Humans had been believed to be safe from the infection of prion diseases of other species for over two hundred years because of the “species barrier,” the relative lack of disease transmission between species (cited by Bruce and colleagues¹¹). However, epidemiological^{4,12} and laboratory studies^{11,13} evidenced the causal association between BSE epidemics and variant Creutzfeldt–Jakob disease (vCJD), the human prion disease.

*Private practice, K-205, Banpodong 929, Sechogu, Seoul, Korea; †Resident, Periodontology and Oral Biology Program, Henry M. Goldman School of Dental Medicine, Boston University; ‡Private practice, 120 South Spalding Drive, Suite 201, Beverly Hills, CA 90212, USA

Corresponding Author: Dr. Hessam Nowzari DDS, PhD, 120 South Spalding Drive, Suite 201, Beverly Hills, CA 90212, USA; email: Hessamnowzari@gmail.com

© 2016 Wiley Periodicals, Inc.

DOI 10.1111/cid.12391

TABLE 1 Types of Creutzfeldt–Jakob Diseases

Type	Cause and clinical features
Kuru	Acquired by cultural cannibalism (legally prohibited in the 1960's). Particular to Papua New Guinea people, prevalence is about 1%. Affects more women and children. Accompanied by tremor, choreiform, and athetoid movements.
Sporadic CJD	Cause Unknown Not common but found worldwide Generally found in people over 50 years old, onset on elder people. Duration <2 years. Short period of illness, ataxia, dementia, myoclonus.
Familial CJD	Genetically caused as result of mutation of PrP gene (PRNP), also called genetic CDJ. Found in approximately 10% of all CJD cases Slightly younger onset than sCJD Longer period of illness: various symptoms dependent on type of mutation.
Iatrogenic CJD	Acquired: transmitted through medical procedures including surgery, transplantation, and blood transfusion. Younger onset than sCJD: ataxia rather than dementia.
Variant CJD	Acquired: assumed to be caused by the consumption of BSE-infected beef (Was initially called new variant CJD [nvCJD]). Duration >6 months. Younger onset: psychiatric features and longer period of illness.

Manufacturers of anorganic bovine bone products claim that they are completely devoid of organic materials, because the presence of organic materials in bovine products indicates the possible existence of pathological prion (PrP^{Sc}) protein, the causative agent of prion diseases. However, plastic surgeons detected proteins including collagens in Bio-Oss[®], Geistlich Pharma AG, Wolhusen, Switzerland, blocks following uneventful patient recovery after orthognathic surgery.¹⁴

Our review of the literature was conceived with the safety of patients in mind. Four questions were put forth for investigation by available literature. The questions asked were those we feel should appropriately be asked and welcomed by manufacturers who prepare bovine bone substitutes. Examination and questioning of these areas should be considered helpful for securing ongoing information that may assist in keeping products safe and the risks to patients as low as possible.

1. Does BSE prion infectivity exist in raw bovine bone?
2. If present, will the infectivity be inactivated by the treatment used for anorganic bovine bone substitutes manufacturing process?
3. Can deproteinization processes remove proteins in anorganic bovine bone substitutes completely?

4. Are current BSE diagnostic tests reliable and valid?

In our paper, we considered the above four questions with references to literature and summarized our search with the caution appropriate to the scientific methods.

Wenz and colleagues reported no protein detection in Bio-Oss[®] and Osteograf/N¹⁵, DENTSPLY Corporate, PA, USA and Sogal and Tofe suggested the risk of BSE transmission from bovine bone graft substitutes should be negligible.¹⁶

However, our systematic review pointed out that their methodologies were erroneous.¹⁷ The Lowry protein assay is used to estimate the content of proteins already in a solution or easily-soluble in dilute alkali,^{18,19} therefore bone matrix should have been extracted and solubilized via defatting and decalcification procedures.²⁰ In addition, prion inactivation should be assessed by bioassay, not by WB because of apparent discrepancies between residual PrP^{Sc} levels in WB and prion inactivation levels.^{21–23}

Although we concluded that bovine-derived graft biomaterials may carry a risk of prion transmission to patients,¹⁷ the risk could not be quantified because of many variables and uncertainties in BSE prion and prion disease. In this paper, additional information will be presented to help understand the risk of BSE prion infection through bovine bone graft materials.

BSE Epidemiology

Control Measures in UK and EU Member States. The origin of BSE remains unknown; however after the first BSE case registered in 1986 epidemiological studies identified contaminated meat and bone meal (MBM) as the most likely vehicle of infection, which led to a series of countermeasures in the United Kingdom (UK).²⁴ In 1988, adding of MBM to ruminants was banned, in 1990 the use of specified bovine offal (SBO; infectious parts such as brain, spinal cord, spleen, and intestines) for all MBM was prohibited,¹⁹ and total ban, prohibiting feeding mammalian protein to any farmed animals, was placed in 1996.²⁵ Approximately 4.1 million cattle were slaughtered between 1996 and 2000 under the Over Thirty Months Scheme (OTMS—the purchase and destruction of all UK cattle over thirty months of age *without testing*).²⁶

Similar control measures were placed in other EU member states at different points after the British BSE outbreak and they were reinforced gradually over time. A total ban was placed throughout the EU in July 2001,²⁵ and EU cofounded OTMS was placed in EU member states for 6 months before the implementation of the active surveillance with rapid testing in July 1, 2001.²⁷

The first case of vCJD in 1996 showed that prions could overcome the species barrier between cattle and humans and research linking the BSE prions being transmitted to humans in UK increased the interest in its pathogenesis.⁴ The results of these efforts began with establishment of the EuroCJD in 1993 and NeuroCJD in 1998 by the European commission.²⁷

Import restrictions against countries with BSE cases were placed in some EU member states in 1996, but trades within the EU community were not mainly affected.²⁸ Conversely, prohibition of the MBM use within a country increased animal product exports to other countries where MBM use was still allowed. When the UK placed the ruminant feed ban in 1988, the export of UK animal feeds doubled the following year, mostly to France. Also, the ruminant feed ban in the EU member states in the early 1990s substantially increased MBM exports from the EU countries to outside the EU, especially into Eastern Europe.^{25,29}

It is claimed that bovine material is obtained from a country with negligible BSE risk, from selected and certified abattoirs, and from the extremity bone

with no BSE infectivity. Our investigation shows that there is no country with negligible BSE risk, contamination of carcasses in abattoirs, and BSE infectivity in various bone tissue.

BSE Surveillance and Epidemiological Trends in UK and Other Affected Countries

Active surveillance was mandated by the EU commission in July 2001.¹⁹ Under the scheme of active surveillance system, animals of target populations (usually, bovines over 30 months of age for the routinely slaughtered healthy population and over 24 months of age for the risk stock) were tested using one of EU-validated 'rapid' tests for abnormal prion protein (PrP^{Sc}), and a positive or an inconclusive case was confirmed by Western blot or immunohistochemistry.³⁰

The size of the BSE epidemic was estimated at 3.5 million for Great Britain³¹ and 0.3 million for France³² using back-calculation modeling based on the data of active surveillance, but the number of BSE cases reported in UK were only 180,000 since 1986.¹⁹

Although it was not operationally implemented in every country, active surveillance detected indigenous BSE cases in 11 countries previously considered as BSE-free (Austria, Czech Republic, Finland, Greece, Italy, Japan, Slovakia, Slovenia, Israel, Poland, Canada, USA, and Sweden).¹⁹ Table 3 shows national prion disease pathology surveillance center cases examined in the United States in 2015.

In addition, atypical forms of BSE were discovered under the system. Since 2003, two types of variants (H-type and L-type) of the classical BSE were detected in several countries including Sweden and the United States with low or unlikely exposure to BSE.³³ Neuropathological and molecular phenotypes of the two atypical BSE differ from each other and the classical BSE. They appeared to occur spontaneously and H-type BSE is associated with mutation E211K within the prion protein gene,³⁴ which is heritable.³⁵

Molecular patterns and brain pathologies of L-type BSE were identical to those of sporadic CJD, a type of human CJD with unknown etiology.^{36,37} Furthermore, serial passages of L-type BSE to non-transgenic mice generated a prion strain similar to that of the classical BSE, implicating the origin of the BSE.³⁸ Taken together, BSE transmission may not be

prevented by control measures such as a meat and bone meal ban.

Many countries do not test cattle for BSE on a regular basis and there are substantial variations in BSE surveillance programs between countries. The EU member states test target populations, while Japan tests all cattle slaughtered for human consumption. Japan detected 31 BSE cases out of 6 million cattle tested between October 2001 and December 2006,³⁹ whereas two US native born cases were detected out of 787. 711 cattle tested during the enhanced surveillance from June 2004 to September 2006.⁴⁰ Considering the numbers of cattle population slaughtered annually (37 million in the US versus 1.26 million in Japan), the proportion of cattle tested in the USA is significantly lower than in Japan.⁴¹

In addition, the age threshold at which cattle had to be tested for BSE has been changing. In January 2009, the age limit of all healthy slaughtered cattle was raised from 30 months to 48 months and in July 2011, from 48 months to 72 months in EU member states including the UK. From January 1, 2013, a minimum sample of healthy slaughtered cattle aged over 72 months will be tested in the UK.⁴²

Most of these control measurements were focused on stopping the spread of the disease to avoid a bigger epidemic event. Prions diseases have a long latency period. The duration varies depending on the type of the disease, but distinction among them is not always clear.^{43–47}

Archived appendix tissues from UK surveys approximate the prevalence of asymptomatic vCJD infection of 1 in 2,000 persons born during 1941 to 1985.⁴⁸ Gill and colleagues suggested that depending on the genetic genotypes some people may harbor the pathologic prion and never develop clinical signs. Therefore, prion transmission through iatrogenic transmission such as blood transfusions, organ transplant, and surgical instrumentation is a constant possibility.^{39,45,48,49}

NeuroPrion was founded in 2003 to facilitate communication between the most important prion research groups, private and public, including 52 research institutes in 20 different countries and federates representing 90% of the public and private research teams in Europe.⁵⁰ (Table 2)

Prion and BSE Diagnostic Tests

Prion. Pathological prion (PrP^{Sc}) is an abnormal isomer of a host-encoded prion protein (PrP^C).³ The

TABLE 2 Current Surveillance Programs

Country	Surveillance program
UK	National CJD Surveillance Unit
EU	European CJD Surveillance Network
Germany	The CJD Surveillance Unit
Argentina	Epidemiologic Surveillance Committee for CJD
USA	National Prion Disease Pathology Surveillance Center
Japan	CJD Surveillance System
Canada	CJD Surveillance Committee
Korea	Center for Infectious Disease Control

prion hypothesis states that PrP^{Sc} is the causative agent of a group of TSE or prion diseases⁵¹ accumulating in individuals affected with most forms of TSE.³

PrP^C was completely degraded with the treatment of 50 µg/mL proteinase K (PK) for 30 minutes at 37°C, whereas PrP^{Sc} from prion-infected Syrian hamster was partially resistant to this treatment, leaving N-terminally truncated 142 amino acid residues (PrP27–30, molecular mass of 27–30 kDa) because of the high β -sheet content.^{51,52}

PrP27–30 and validity of Current BSE Diagnostic Tests. PrP27–30, amino acid residues of PrP^{Sc} after PK treatment, has been used as a surrogate marker for prion infectivity and most TSE diagnostic tests currently available rely on detection of PrP27–30. However, a number of studies have shown that prion infectivity does not always correlate with the presence of PrP^{Sc} or PK27–30. A recent study by Balkema-Buschmann and colleagues, indicates there is no concurrence between PrP^{Sc} detection and BSE infectivity.⁵³ None of the TSE diagnostic tests used (IHC, SAF immunoblot, and PMCA) detected PrP^{Sc} in peripheral nerves, tongue, and nasal mucosa, while 30 to 92% of transgenic mice developed the disease after tissues homogenates inoculation.

In Creutzfeldt–Jakob disease (CJD)-infected mice, PK27–30 levels of CNS microglia were 50 fold less than those of undiluted brain homogenates with WB, but the infectivity titers of the two tissue types were similar to each other.⁵⁴ None or only traces of PK27–30 were detected in 263K scrapie infected hamster brain fractions containing high infection titers⁵⁵ and

TABLE 3 National Prion Disease Pathology Surveillance Center Cases Examined in US* (August 25, 2015)

Year	Prion disease	Sporadic CJD	Familial CJD	Iatrogenic CJD	Variant CJD
2000	101	89	12	0	0
2001	118	110	8	0	0
2002	144	125	17	2	0
2003	159	138	21	2	0
2004	180	162	17	0	1 [†]
2005	179	157	21	1	0
2006	182	163	17	1	2 [‡]
2007	206	187	19	0	0
2008	223	207	16	0	0
2009	233	212	20	1	0
2010	247	218	29	0	0
2011	239	216	23	0	0
2012	241	218	23	0	0
2013	257	222	34	1	0
2014	210	187	21	1	1 [§]
2015	137	118	8	0	0
TOTAL	3056 [¶]	2729	306 ^{**}	9	4

*Listed based on the year of death or, if not available, on year of referral

[†]Disease acquired in the United Kingdom

[‡]Disease acquired in the United Kingdom in one case and in Saudi Arabia in the other.

[§]Disease possibly acquired in a Middle Eastern or Eastern European country

[¶]Includes 12 (11 from 2015) cases with type determination pending in which the diagnosis of vCJD has been excluded. The sporadic cases include 2853 cases of sporadic Creutzfeldt-Jakob disease (sCJD), 54 cases of Variably Protease-Sensitive Prionopathy (VPSPr) and 22 cases of sporadic Fatal Insomnia (sFI).

**Total excludes 194 familial blood only cases.

PrP^{Sc} levels were extremely low or undetectable by WB, IHC, CDI, or immunoprecipitation in either hamster 263K scrapie or human GSS (Gerstmann Sträussler Scheinker; a type of human TSE) disease-infected transgenic mice brain tissues containing high infectivity titers.⁵⁶

Conversely, PK-sensitive PrP^{Sc} has been discovered in prion-infected humans and animals^{57,58} and abnormal PrP^{Sc} (~6 kDa), which was not detected with standard diagnostic procedures was also discovered in a novel human prion disease.^{59,60} Furthermore, several studies showed discrepancies in tissue PrP^{Sc} distribution when different diagnostic tests were used, which may be attributed to uneven PrP^{Sc} distribution within tissues or organs,⁶¹ different diagnostic

sensitivity between tests,^{60,62} relatively low PrP^{Sc} accumulation in non-CNS tissues in cattle, and so forth. Conflicting results were also found in some CNS tissue samples of spinal cord and rostral medulla. In addition, when different antibodies (R145; F99) were applied for IHC, the test results differed in multiple samples of trigeminal ganglion and dorsal root ganglion.⁶³

The exact portion and the size of brain samples for BSE diagnostic tests or the antibody for IHC may vary among laboratories. However, inconsistent PrP^{Sc} detection test results may be problematic in the context of the BSE confirmatory tests because WB or IHC are used to confirm a positive or an inconclusive case with a rapid test for the large screening.⁶⁴

BSE Infectivity in Bovine Bone Before and After Manufacturing Processing. BSE infectivity and PrP^{Sc} were detected in sternal bone marrow in experimental and serum samples from BSE-confirmed cases using flow cytometry.⁶⁵ In spite of BSE regulations such as prohibition of the use of particular types of stunning methods and SRM (specified risk materials that most likely contain BSE agents such as brain, spinal cord, intestine, tonsil, etc.) removal, carcass contamination with CNS tissue occurred at slaughter houses and cutting plants. For example, spinal cord removal prior to carcass splitting reduced the carcass contamination level but could not prevent carcass contamination.^{66,67} Under the simulated commercial abattoir condition, less than 1.5 mg (maximum 9 mg) spinal cord tissue from a previous carcass was recovered in four subsequent carcasses.⁶⁸ Contamination or cross-contamination of bovine carcasses by CNS tissue, which frequently occurred in abattoirs, was not removed by various washing procedures.⁶⁷

Proteins including collagens were detected in some anorganic bovine bone substitutes including Tutoplast[®], Zimmer Dental, CA, USA (bovine), Bio-Oss[®], and tibia samples treated at the similar condition for Bio-Oss[®] deproteinization process,^{68–70} and most importantly, whether BSE prion is inactivated by anorganic bovine bone manufacturing processes has not been proven yet.¹⁷

Prions are well known for their resistance to conventional chemical and physical decontamination methods, and the heat treatment used for anorganic bovine bone material preparation (300°C for Bio-

Oss[®], 1100°C for PepGen P-15[®], DENTSPLY Corporate, PA, USA)⁶⁹ has not proven to inactivate BSE prion. Brain tissue from scrapie-infected hamsters transmitted the disease after exposure to dry heat at 600°C for 15 minutes, but no transmission at 1000°C for 5 min.^{70,71} Whether BSE prion has similar resistance to inactivation as scrapie prion at the same heat treatment condition is unknown.

CONCLUSION

The limited ability to screen prions within the animal genome along with a long latency period to manifestation of the disease (1 to over 50 years) in infected patients provides a framework for discussing possible long-term risks of the xenografts that are used so extensively in dentistry. Patient counseling should include a clear description of the origin of the graft. A close evaluation of published papers in favor of bovine bone reveals wide variation in study designs, creating conditions for erroneous conclusions from the methods used. Often, use of broad or vague definitions and errors in analytical models can be noted. With the advancements made in bone biology, the available alternatives should be considered. We suggest abolishing the use of the bovine bone.

ACKNOWLEDGMENT

The authors would like to thank the National Prion Disease Pathology Surveillance Center (NPDPS) in Cleveland, Ohio, for the data shared.

CONFLICT OF INTEREST STATEMENT

No conflict of interest.

REFERENCES

1. Baadsgaard K. Kiel bone in the treatment of pseudarthrosis. An experimental study. *Acta Orthop Scand* 1969; 40: 696–707 (Orell S 1934)
2. Campana V, Milano G, Pagano E, et al. Bone substitutes in orthopaedic surgery: from basic science to clinical practice. *J Mater Sci Mater Med* 2014; 25:2445–2461.
3. Aguzzi A, Heikenwalder M, Miele G. Progress and problems in the biology, diagnostics, and therapeutics of prion diseases. *J Clin Invest* 2004; 114:153–160.
4. Will RG, Ironside JW, Zeidler M, et al. A new variant of Creutzfeldt–Jakob disease in the UK. *Lancet* 1996; 347: 921–925.
5. Collinge J. Prion diseases of humans and animals: their causes and molecular basis. *Annu Rev Neurosci* 2001; 24: 519–550.
6. Collins S, Boyd A, Lee JS, et al. Creutzfeldt–Jakob disease in Australia 1970–1999. *Neurology* 2002; 59:1365–1371.
7. Creutzfeldt H. Übereineigenartigeherdförmige Erkrankung des zentralnervensystems (Vorläufige Mitteilung). *Zeitschrift für die gesamte Neurologie und Psychiatrie* 1920; 57:1–118.
8. Jakob A. Übereineigenartige Erkrankungen des zentralnervensystems mit bemerkenswerten anatomischen Befunden. *Zeitschrift für die gesamte Neurologie und Psychiatrie* 1921; 64:147–228.
9. Aguzzi A, Sigurdson C, Heikenwalder M. Molecular mechanisms of prion pathogenesis. *Annu Rev Pathol* 2008; 3:11–40; PubMed; PMID:18233951; <http://dx.doi.org/10.1146/annurev.pathmechdis.3.121806.154326>
10. Prusiner SB. Cell biology. A unifying role for prions in neurodegenerative diseases. *Science* 2012; 336:1511–3. <http://dx.doi.org/10.1126/science.1222951>
11. Bruce M, Chree A, McConnell I, Foster J, Pearson G, Fraser H. Transmission of bovine spongiform encephalopathy and scrapie to mice: strain variation and the species barrier. *Philos Trans R Soc Lond B Biol Sci* 1994; 343:405–411. Review
12. Wilesmith JW, Ryan JB, Arnold ME, Stevenson MA, Burke PJ. Descriptive epidemiological features of cases of bovine spongiform encephalopathy born after July 31, 1996 in Great Britain. *Vet Rec* 2010; 167:279–286.
13. Collinge J, Sidle KC, Meads J, Ironside J, Hill AF. Molecular analysis of prion strain variation and the aetiology of ‘new variant’ CJD. *Nature* 1996; 383:685–690.
14. Hönlig JF, Merten HA, Heinemann DE. Risk of transmission of agents associated with Creutzfeldt–Jakob disease and bovine spongiform encephalopathy. *Plast Reconstr Surg* 1999; 103:1324–1325.
15. Sogal A, Tofe AJ. Risk assessment of bovine spongiform encephalopathy transmission through bone graft material derived from bovine bone used for dental applications. *J Periodontol* 1999; 70:1053–1063.
16. Wenz B, Oesch B, Horst M. Analysis of the risk of transmitting bovine spongiform encephalopathy through bone grafts derived from bovine bone. *Biomaterials* 2001; 22: 1599–606.
17. Kim Y, Nowzari H, Rich SK. Risk of prion disease transmission through Bovine-Derived bone substitutes: a systematic review. *Clin Implant Dent Relat Res* 2013; 15: 645–653.
18. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951; 193:265–275.

19. Ducrot C, Arnold M, de Koeijer A, Heim D, Calavas D. Review on the epidemiology and dynamics of BSE epidemics. *Vet Res* 2008; 39:15. Epub 2008 Jan 11. Review.
20. Anastassiades T, Puzic O, Puzic R. Effect of solubilized bone matrix components on cultured fibroblasts derived from neonatal rat tissues. *Calcif Tissue Res* 1978; 26:173–179.
21. Giles K, Glidden DV, Beckwith R, et al. Resistance of bovine spongiform encephalopathy (BSE) prions to inactivation. *PLoS Pathog* 2008; 4:e1000206. Epub 2008 Nov 14.
22. McLeod AH, Murdoch H, Dickinson J, et al. Proteolytic inactivation of the bovine spongiform encephalopathy agent. *Biochem Biophys Res Commun* 2004; 317:1165–1170. Erratum in: *Biochem Biophys Res Commun* 2004; 321:1069.
23. Peretz D, Supattapone S, Giles K, et al. Inactivation of prions by acidic sodium dodecyl sulfate. *J Virol* 2006; 80:322–331.
24. Barria MA, Ironside JW, Head MW. Exploring the zoonotic potential of animal prion diseases: in vivo and in vitro approaches. *Prion* 2014; 8:85–91.
25. Ducrot C, Sala C, Ru G, et al. Modelling BSE trend over time in Europe, a risk assessment perspective. *Eur J Epidemiol* 2010; 25:411–419. Epub 2010 Apr 13.
26. FSA review of BSE control 2000, World Health Organization, Edited by Carlos Dora.
27. EURO-CJD. Genetic epidemiology of Creutzfeldt–Jakob disease in Europe. *Rev Neurol (Paris)* 2001; 157:633–637.
28. Heim D, Gardner I, Mumford E, Kihm U. Risk assessment and surveillance for bovine spongiform encephalopathy. *Rev Sci Tech* 2006; 25:937–950. Review.
29. Butler D. Did UK 'dump' contaminated feed after ban? *Nature* 1996; 381:544–545.
30. Grassi J, Maillet S, Simon S, Morel N. Progress and limits of TSE diagnostic tools. *Vet Res* 2008; 39:33. Epub 2008 Feb 15. Review.
31. Donnelly CA, Ferguson NM, Ghani AC, Anderson RM. Implications of BSE infection screening data for the scale of the British BSE epidemic and current European infection levels. *Proc Biol Sci* 2002; 269:2179–2190.
32. Supervie V, Costagliola D. The unrecognized French BSE epidemic. *Vet Res* 2004; 35:349–362.
33. Stack MJ, Focosi-Snyman R, Cawthraw S, Davis L, Chaplin MJ, Burke PJ. Third atypical BSE case in Great Britain with an H-type molecular profile. *Vet Rec* 2009; 165:605–606.
34. Richt JA, Hall SM. BSE case associated with prion protein gene mutation. *PLoS Pathog* 2008; 4:e1000156.
35. Nicholson EM, Brunelle BW, Richt JA, Kehrl ME Jr, Greenlee JJ. Identification of a heritable polymorphism in bovine PRNP associated with genetic transmissible spongiform encephalopathy: evidence of heritable BSE. *PLoS One* 2008; 3:e2912.
36. Casalone C, Zanusso G, Acutis P, Ferrari S, Capucci L, Tagliavini F, Monaco S, Caramelli M. Identification of a second bovine amyloidotic spongiform encephalopathy: molecular similarities with sporadic Creutzfeldt–Jakob disease. *Proc Natl Acad Sci USA* 2004; 101:3065–3070. Epub 2004 Feb 17.
37. Comoy EE, Casalone C, Lescoutra-Etcheagaray N, et al. Atypical BSE (BASE) transmitted from asymptomatic aging cattle to a primate. *PLoS One* 2008; 3:e3017.
38. Capobianco R, Casalone C, Suardi S, et al. Conversion of the BASE prion strain into the BSE strain: the origin of BSE? *PLoS Pathog* 2007; 3:e31.
39. Kadohira M, Stevenson MA, Kanayama T, Morris RS. Epidemiology of bovine spongiform encephalopathy in cattle in Hokkaido, Japan, between September 2001 and December 2006. *Vet Rec* 2008; 163:709–713.
40. Federal Register. Rules and Regulations, 2008; 73, <http://www.fda.gov/OHRMS/DOCKETS/98fr/08-1180.pdf> (Accessed April 25, 2008).
41. Brink S, Shute N. Is it safe? New beef rules aim to stop mad cow disease. But they may not be enough—and there's too much we don't know. *US News World Rep* 2004; 136:16–21.
42. United Kingdom Department of Agriculture and Rural Development. www.dardni.gov.uk/change-to-bse-testing.doc (Accessed March 19, 2011).
43. Barash J. Identification of Creutzfeldt–Jakob disease variants. *Arch Neurol* 2009; 66:1045 [author reply 1045–1046]; PubMed <http://dx.doi.org/10.1001/archneurol.2009.143>
44. Diack AB, Head MW, McCutcheon S, et al. Variant CJD: 18 years of research and surveillance. *Prion* 2014; 8:286–295; PubMed <http://dx.doi.org/10.4161/pri.29237>
45. Maheshwari A, Fischer M, Gambetti P, et al. Recent US Case of Variant Creutzfeldt–Jakob Disease—Global Implications. *Emerg Infect Dis* 2015; 21:750–759. doi:10.3201/eid2105.142017.
46. Zerr I, Kallenberg K, Summers DM, et al. Updated clinical diagnostic criteria for sporadic Creutzfeldt–Jakob disease. *Brain* 2009; 132:2659–2668.
47. Ugnon-Café S, Dorey A, Bilheude JM, et al. Rapid screening and confirmatory methods for biochemical diagnosis of human prion disease. *J Virol Methods* 2011; 175:216–223; PubMed <http://dx.doi.org/10.1016/j.jviromet.2011.05.016>
48. Gill ON, Spencer Y, Richard-Loendt A, et al. Prevalent abnormal prion protein in human appendixes after bovine spongiform encephalopathy epizootic: largescale survey. *BMJ* 2013; 347:f5675. <http://dx.doi.org/10.1136/bmj.f5675>

49. Thomas JG, Chenoweth CE, Sullivan SE. Iatrogenic Creutzfeldt-Jakob disease via surgical instruments. *J Clin Neurosci* 2013; 20:1207–1212.
50. <http://www.neuropion.org/en/home.html>
51. Pan KM, Baldwin M, Nguyen J, et al. Conversion of alpha-helices into beta-sheets features in the formation of the scrapie prion proteins. *Proc Natl Acad Sci USA* 1993; 90:10962–10966.
52. Prusiner SB. Prions. *Proc Natl Acad Sci USA* 1998; 95: 13363–13383. Review. (Nobel lecture)
53. Balkema-Buschmann A, Eiden M, Hoffmann C, et al. BSE infectivity in the absence of detectable PrPSc accumulation in the tongue and nasal mucosa of terminally diseased cattle. *J Gen Virol* 2011; 92(Pt 2):467–476. Epub 2010 Oct 13.
54. Baker CA, Martin D, Manuelidis L. Microglia from Creutzfeldt-Jakob disease-infected brains are infectious and show specific mRNA activation profiles. *J Virol* 2002; 76:10905–10913.
55. Berardi VA, Cardone F, Valanzano A, Lu M, Pocchiari M. Preparation of soluble infectious samples from scrapie-infected brain: a new tool to study the clearance of transmissible spongiform encephalopathy agents during plasma fractionation. *Transfusion* 2006; 46:652–658.
56. Barron RM, Campbell SL, King D, Bellon A, Chapman KE, Williamson RA, Manson JC. High titers of transmissible spongiform encephalopathy infectivity associated with extremely low levels of PrPSc in vivo. *J Biol Chem* 2007; 282: 35878–35886. Epub 2007 Oct 8.
57. Safar JG, Geschwind MD, Deering C, et al. Diagnosis of human prion disease. *Proc Natl Acad Sci USA* 2005; 102: 3501–3506.
58. Thackray AM, Hopkins L, Bujdoso R. Proteinase K-sensitive disease-associated ovine prion protein revealed by conformation-dependent immunoassay. *Biochem J* 2007; 401:475–483.
59. Gambetti P, Dong Z, Yuan J, et al. A novel human disease with abnormal prion protein sensitive to protease. *Ann Neurol* 2008; 63:697–708.
60. Head MW, Knight R, Zeidler M, Yull H, Barlow A, Ironside JW. A case of protease sensitive prionopathy in a patient in the UK. *Neuropathol Appl Neurobiol* 2009; 35: 628–632. Epub 2009 Aug 7.
61. Safar JG, Scott M, Monaghan J, et al. Measuring prions causing bovine spongiform encephalopathy or chronic wasting disease by immunoassays and transgenic mice. *Nat Biotechnol* 2002; 20:1147–1150. Epub 2002 Oct 21.
62. Soto C, Anderes L, Suardi S, et al. Pre-symptomatic detection of prions by cyclic amplification of protein misfolding. *FEBS Lett* 2005; 579:638–642.
63. Arnold ME, Ryan JB, Konold T, et al. Estimating the temporal relationship between PrPSc detection and incubation period in experimental bovine spongiform encephalopathy of cattle. *J Gen Virol* 2007; 88(Pt 11): 3198–3208.
64. Trieschmann L, Navarrete Santos A, Kaschig K, Torkler S, Maas E, Schätzl H, Böhm G. Ultra-sensitive detection of prion protein fibrils by flow cytometry in blood from cattle affected with bovine spongiform encephalopathy. *BMC Biotechnol* 2005; 5:26.
65. Helps CR, Hindell P, Hillman TJ, et al. Contamination of beef carcasses by spinal cord tissue during splitting. *Food Control* 2002; 13:417–423.
66. Takada N, Horiuchi M, Sata T, Sawada Y. Evaluation of methods for removing central nervous system tissue contamination from the surface of beef carcasses after splitting. *J Vet Med Sci* 2008; 70:1225–1230.
67. Helps CR, Fisher AV, Harbour DA, O'Neill DH, Knight AC. Transfer of spinal cord material to subsequent bovine carcasses at splitting. *J Food Prot* 2004; 67:1921–1926
68. Tadic D, Epple M. A thorough physicochemical characterisation of 14 calcium phosphate-based bone substitution materials in comparison to natural bone. *Biomaterials* 2004; 25:987–994.
69. Brown P, Rau EH, Lemieux P, Johnson BK, Bacote AE, Gajdusek DC. Infectivity studies of both ash and air emissions from simulated incineration of scrapie-contaminated tissues. *Environ Sci Technol* 2004; 38:6155–60.
70. Brown P. BSE and transmission through blood. *Lancet* 2000; 356:955–956.
71. Lee J, Kim SY, Hwang KJ, Ju YR, Woo HJ. Prion diseases as transmissible zoonotic diseases. *Osong Public Health Res Perspect* 2013; 4:57–66.